RESEARCH ARTICLE

IN VITRO PROPAGATION OF HOLOSTEMMA ADA-KODIEN SCHULT. AN ENDANGERED MEDICINAL PLANT

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ABSTRACT

Holostemma ada-kodien is a medicinal plant belongs to the family Asclepiadaceae. Root tubers contain alpha amyrin, Lupeol, and β- Sitosterol. The present study was to standardize in vitro culture of Holostemma ada-kodien Schult. On Murashige and Skoog (MS) medium supplemented with different combinations of growth regulators to induce shoot, root and callus formation. Among several combinations of BAP, NAA and Kinetin studied, nutrient medium supplemented with 0.1mg/l BAP and 1.0 mg/l Kinetin induced maximum number of shoots. Highest callus proliferation was observed in the medium supplemented with 1.0 mg/l BAP and 1.0 mg/l NAA. Root initiation was also observed in the same medium.

**Key words:** Multiple shoots, callus proliferation, root initiation, Holostemma ada-kodien.

INTRODUCTION

*Holostemma ada-kodien* is a medicinal plant belongs to the family Asclepiadaceae. Tuberous roots of the plant are medicinally important and are utilized as a major ingredient of the drug “jivanthi” (Kolammal, 1979). The plant is a twining shrub with opposite leaves, and roots are developed in axillary umbellate cymes. Roots contain α-amyrin, Lupeol and β- Sitosterol. The roots are used for cough, fever, ophthalmic diseases, stomachache, spermatorrhoea, emaciation, dysentery, tuberculosis, arrested urination, scorpion bite, kidney stones, goiter etc. About 150 metric tons of root tubers per annum are required to major southern Indian pharmacies for Ayurvedic preparations (Nair et al. 1992). The destructive and merciless collection of tubers, in recent times, has led to extreme loss of the plant in their natural habitats and consequently it is listed out as vulnerable and rare in FRLHT red list (FRLHT, 1997). Conventionally propagated through seeds, stem cutting and root cuttings is too slow to meet the demand of this invaluable therapeutic plant. Hence, in vitro tissue culture method might be of great value as an alternate method to achieve rapid multiplication of this species. In the last few decades, plant tissue culture has played an important role for production of large number of plants, independent of seasons, aiming at re-establishment of these plants in their natural habitat. Besides, they also serve as an alternate source of plant raw materials for commercial utilization, which decreases pressure on the plant population in wild habitat (Krishnan et al., 2011). Micropropagation through axillary bud multiplication (Sudha et al., 1998) and indirect organogenesis from callus (Sudha et al., 2000) has been reported on this species. An efficient protocol for rapid propagation of *H. ada-kodien* by using shoot tip and nodal explants has been developed in the present study.

MATERIALS AND METHODS

Mother plants of *Holostemma ada-kodien* were collected from Centre for Medicinal Plants Research (CMPR), AryaVaidya Sala, Changuvetty, Kottakkal. The explants were initially washed in running tap water for about 30 minutes, then with detergent extran for about 5-10 minutes and finally, they were thoroughly washed in distilled water. The explants were surface sterilized in 0.1% mercuric chloride for 10 minutes, followed by thorough washing in distilled water. Then the explants were cut into appropriate size and cultured on MS basal medium. After 30 days, shoots formed were sub-cultured on MS medium containing various combinations of BAP, NAA and Kinetin.

RESULTS AND DISCUSSION

Culture initiation

1-2 shoots were formed from nodal explants when cultured on MS basal medium. After 30 days, shoots formed were sub-cultured on MS media containing various combinations of BAP, NAA and Kinetin Table (1).

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Holostemma ada-kodien initiated in M S medium containing 0.1mg/l BAP and 1.0mg/l Kinetin. High callus proliferation is also observed.

Axillary Bud Multiplication

Axillary bud proliferation was observed from sub-cultured nodal explants when cultured on MS medium containing different combinations of PGRs. About 4 shoots/explant were induced in medium containing Kinetin alone, whereas BAP alone induced about 3 shoots/explant. BAP in combination with Kinetin induced multiple shoots from axillary bud. Medium containing 0.1mg/l BAP and 1.0 mg/l Kinetin produced highest number of shoots (10 shoots/explant) whereas the reverse combination facilitated the least number of axillary buds. About 6 shoots/explant were observed in the medium containing 1.0mg/l kinetin in combination with 0.5mg/l NAA. According to Sudha et al. (1998) 1.0 mg l–1 BAP in combination with 0.1 mg l–1 NAA was effective, and they obtained 3.8 shoots per node. Callus proliferation was low in this combination whereas the medium having 1.0mg/l BAP and 0.5mg/l NAA induced only about 2 shoots/explant with high callus proliferation. The present study provides information about the efficacy of Kinetin in combination with low concentration of BAP in facilitating axillary bud multiplication. This is in accordance with the view that Holostemma ada-kodien has high indigenous concentration of auxin. Thus, axillary bud proliferation in H. ada-kodien is more depended on the concentration of which cytokinin is used. High proliferation of axillary buds was observed after trimming off the initial axillary shoots from the main shoot. Among the different cultures medium containing 1.0 mg/l Kinetin with 0.5mg/l NAA showed such rapid proliferation of axillary buds after trimming. This also shows the effect of kinetin in the rapid proliferation of H.ada-kodien.

The described protocol could be used for the rapid propagation, conservation and genetic transformation studies of this

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<th>Growth regulators(mg/l)</th>
<th>Number of shoots/node</th>
<th>Callus proliferation</th>
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<tr>
<td>BAP</td>
<td>NAA</td>
<td>Kinetin</td>
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<td>Hormone free</td>
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Table 1. Effect of MS media provided with different combinations of growth regulators on shoot and callus multiplication of H ada-kodien. Growth period was 30days.
valuable, endangered medicinal plant through axillary bud multiplication.

REFERENCES

FRLHT, 1997. Medicinal plants of India- Guidelines for national policy and conservation programs (Foundation for Revitalization of Local Health Traditions), Bangalore (India).


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